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## Total Synthesis and Absolute Configuration of Macrocidin A, a Cyclophane Tetramic Acid Natural Product\*\*

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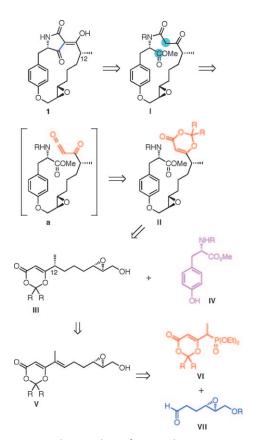
Macrocidin A (1) and macrocidin B (2) represent a new family of plant pathogens produced by *Phoma macrostoma*, a microorganism parasitic to Canadian thistle.[1] The intriguing structure of 1, which includes a tetramic acid<sup>[2]</sup> group installed in a cyclophane skeleton, was determined by extensive 2D NMR studies and single-crystal X-ray analysis, although the absolute configuration remains to be addressed because of a paucity of the natural sample. The macrocidins have significant herbicidal activity on broadleaf weeds but not on grasses, which makes them a potential lead for new herbicide design. Their biological activity and novel chemical structures have made these compounds attractive targets for chemical synthesis. Whilst construction of the macrocyclic skeleton has been addressed, [3] the synthesis of the full structure remains to be achieved. Herein, we describe the first total synthesis of macrocidin A (1) using macrolactam formation followed by cyclization.

Scheme 1 outlines our retrosynthetic analysis based upon the construction of the acyltetramic acid moiety using the Lacey–Dieckmann cyclization<sup>[4]</sup> of macrolactam **I**, which in turn would be accessible by the intramolecular trapping of an acylketene species (a) that may be thermally generated from dioxinone precursor **II**.<sup>[5]</sup> This key intermediate (**II**) could be

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Scheme 1. Retrosynthetic analysis of macrocidin A.

assembled from the stereodefined epoxy alcohol **III** and tyrosine unit **IV**. One of the challenges in the synthesis of **III** was the establishment of the C12 stereogenic center, <sup>[6]</sup> for which we planned to employ either a substrate- or catalyst-controlled diastereoselective hydrogenation of trisubstituted olefin **V**. Finally, olefin **V** could be obtained from phosphonate **VI** and aldehyde **VII**.

Scheme 2 shows the preparation of trisubstituted olefins **10** and **11**, which began with the two-step conversion of propargyl alcohol into allyl alcohol **5**.<sup>[7]</sup> Epoxide **6**, which was prepared by a Katsuki–Sharpless asymmetric epoxidation reaction (93 % ee), [8] was silylated and subjected to rhodium-catalyzed hydroboration, [9] followed by oxidation to give alcohol **8**. Swern oxidation of **8**, followed by a Horner–Emmons reaction with chiral phosphonate **12**<sup>[10]</sup> afforded olefin **10** (E/Z=9:1). Recrystallization (n-hexane/ethyl acetate) gave pure (E)-**10**, which had a stereodefined trisubstituted olefin and a menthone chiral auxiliary<sup>[11]</sup> ready for diastereoselective hydrogenation. The substrate for the catalyst-controlled diastereoselective hydrogenation, aceto-

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Scheme 2. Preparation of substrates 10 and 11. a) Allyl bromide, Cul, NaI, K2CO3, acetone, RT, 5 h (85%). b) LiAlH4, THF, reflux, 2 h (75%). c) (iPrO)<sub>4</sub>Ti, L-(+)-diethyl tartrate, tBuOOH, M.S. 4 Å, CH<sub>2</sub>Cl<sub>2</sub>, -20°C, 24 h (80%, 93% ee). d) tBuPh<sub>2</sub>SiCl, imidazole, DMF, RT, 1 h (99%). e) Catecholborane, [Rh(PPh<sub>3</sub>)<sub>3</sub>Cl], THF, 0°C, 1.5 h; H<sub>2</sub>O<sub>2</sub>, pH 7 phosphate buffer (72%). f) (COCl)2, DMSO, Et3N,  $CH_2Cl_2$ ,  $-78 \rightarrow 0$  °C, 1.5 h, (quant.). g) **12**, LiN(*i*Pr)<sub>2</sub>, THF,  $-78 \rightarrow 0$  °C (99%, E/Z = 9:1); recrystallization (n-hexane/EtOAc), 57%. h) 13, LiN(iPr)2, HMPA, THF,  $-78 \rightarrow 0$  °C (96%, E/Z = 9:1), SiO<sub>2</sub> gel chromatography. M.S. = molecular sieves, TBDPS = tert-butyldiphenylsilyl, DMF = N,N-dimethylformamide, DMSO = dimethyl sulfoxide, HMPA = hexamethylphosphoramide

nide 11, was also prepared in a similar manner, except for the use of HMPA as a co-solvent in the olefination step to prevent aggregation of the phosphonate anion that is derived from 13. The E/Z ratio was again 9:1, but (E)-11 was easily isolated by silica gel column chromatography.

In addressing the controlled construction of the C12 stereogenic center, our initial attempts used the chiral, nonracemic substrate 10, hoping that the menthone moiety would control the diastereomeric facial selection (Scheme 3). However, no reaction occurred with homogeneous catalysts, such as Wilkinson<sup>[12a]</sup> or Crabtree<sup>[12b]</sup> complexes, presumably owing to the high steric hindrance around the olefin. Conversely, several heterogeneous catalysts were active enough to give

Scheme 3. Diastereoselective hydrogenation of 10.

the product (14) in excellent yield, although the stereoselectivity was uniformly 1:1. Therefore, unfortunately, the stereoselective influence of the chiral auxiliary on the hydrogenation reaction was negligible.

At this stage, we decided to investigate the catalystcontrolled asymmetric hydrogenation reaction, primarily considering iridium catalysts with chiral N,P ligands that are known to be uniquely effective for unfunctionalized olefins and do not require neighboring group participation. [13] Attempts at the enantioselective hydrogenation of prochiral substrate 15 with catalysts A-E (Scheme 4) were mostly

OSi(
$$Pr$$
)<sub>3</sub>

A (2 mol%)

CF<sub>3</sub>CH<sub>2</sub>OH

16

conditions

RT, 12 h

18% (69% recovery of 15)

40 °C, 12 h

40% (38% recovery of 15)

catalysts

CF<sub>3</sub>

A

CF<sub>3</sub>

Scheme 4. The attempted asymmetric hydrogenation of 15.

disappointing, with poor catalytic activities observed except in the case of catalyst A.<sup>[13a]</sup> In the presence of 2 mol % of A(CF<sub>3</sub>CH<sub>2</sub>OH; room temperature; 12 h), the highly enantioselective hydrogenation of 15 did proceed to give the desired product 16 with excellent stereoselectivity (97:3), albeit in low yield (18%). The problem was the low reactivity, and even at pressures as high as 10 MPa the reaction was sluggish. When carried out at 40 °C, a slightly higher yield of 16 was obtained (40%) with recovery of **15** in 38% yield. However, no further improvement was possible, either by extending the reaction time or by increasing the catalyst loading, because the epoxide moiety seemed to be damaged by the fairly high Lewis acidity of the iridium catalyst or the Brønsted acidity of iridium hydride intermediates.

After considerable experimentation, a solution was found by converting the epoxide into the corresponding iodohydrin prior to the hydrogenation (Scheme 5). Therefore, alcohol 17 was prepared by the desilylation of 11 (nBu<sub>4</sub>NF, THF, 0°C, 1 h, 99%), which allowed the highly regioselective (92:8) epoxide ring opening (NaI, B(OAc)<sub>3</sub>, AcOH, acetone),<sup>[14]</sup> to give 1,3-diol 18 as the major product along with a small amount of 1,2-diol 18' in 94% combined yield (18/18' = 92:8); these products were easily separated by silica gel column

Scheme 5. Hydrogenation of 18. a) NaI, B(OAc)3, AcOH, acetone,  $-20\rightarrow0$  °C, 2 h (94%). b) H<sub>2</sub> (10 MPa), **A** (2 mol%), CF<sub>3</sub>CH<sub>2</sub>OH, 40°C, 12 h (96%). c) K<sub>2</sub>CO<sub>3</sub>, methanol, RT, 1 h (quant.).

chromatography.<sup>[15]</sup> Pleasingly, the attempted hydrogenation of 18 in the presence of iridium catalyst A proceeded smoothly, without any reductive fission of the C-I bond, [16] affording the desired compound 19 in excellent yield (96%) and excellent stereoselectivity (97:3).[17] Treatment of 19 with potassium carbonate in methanol regenerated the oxirane ring to regioselectively afford epoxy alcohol 20 in quantitative vield.[18]

For preparation of the macrolactam precursor, tyrosine unit 21[19] was coupled with epoxy alcohol 20 using the Mitsunobu reaction (TMAD and nBu<sub>3</sub>P);<sup>[20]</sup> detachment of the benzyloxycarbonyl protecting group then gave amine 23. The pivotal construction of the macrocidin skeleton involved two stages: 1) macrolactam formation via intramolecular ketene trapping, and 2) Lacey-Dieckmann cyclization for constructing the tetramic acid (Scheme 6).

Scheme 6. Attempts for the macrolactam formation and the Lacey-Dieckmann condensation. a) TMAD, nBu<sub>3</sub>P, toluene, RT, 3 h (89%). b) H<sub>2</sub>, Pd(OH)<sub>2</sub>/C, MeOH, RT, 4 h (98%). c) Toluene, reflux, 2 h (90%). Cbz = benzyloxycarbonyl, TMAD = N, N, N', N'-tetramethyl azodicarboxamide.

Indeed, we were pleased to find that the first step was realized quite smoothly by the heating of 23 in toluene (reflux, 2 h), giving macrolactam 24 in 90% yield, ready for the final step of the total synthesis. However, we were disappointed by the complete failure of the attempted Lacey-Dieckmann condensation to convert macrolactam 24 into macrocidin A (1) using tBuOK or various other basic reagents (e.g.  $nBu_4NF$ , NaOMe).

Single-crystal X-ray diffraction analysis of 24<sup>[21]</sup> suggested an inherent problem in the cyclization substrate: the distance between C10 and C19 $^{[6]}$  was too large (4.61 Å) to allow bond formation (Figure 1). To bring the two centers closer together,

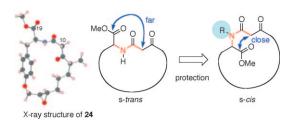


Figure 1. Conformational analysis of macrolactam 24 and a strategy for the construction of the tetramic acid ring.

the geometry of the amide group would have to be changed from the s-trans to the s-cis conformation. It occurred to us that such a conformational change would be achieved if a protecting group was introduced at the amide nitrogen atom, thus providing a more favorable situation for the cyclization.

Along these lines, we prepared para-azidobenzyl (PAB)<sup>[22]</sup> derivative 28, which was later employed in the eventual route, starting from the Mitsunobu reaction (DEAD, PPh<sub>3</sub>) of epoxy alcohol 20 and phenol 25[23] (Scheme 7). In this instance, the

Scheme 7. Synthesis of para-azidobenzyl-protected macrocidin A (29). a) DEAD, PPh3, toluene, RT, 3 h (89%). b) nBu4NF, THF, RT, 4 h (91%). c) Toluene, reflux, 2 h (86%). d) tBuOK, tBuOH, THF, RT, 30 min (87%). DEAD = diethyl azodicarboxylate, Teoc = 2-(trimethylsilyl) ethoxycarbonyl. PAB = p-azidobenzyl.

use of TMAD and nBu<sub>3</sub>P was not suitable, because the azide moiety in phenol 25 was reduced by the latter reagent. The 2-(trimethylsilyl)ethoxycarbonyl (Teoc) group in 26 was

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removed to give compound **27**, which smoothly underwent macrolactam formation to **28** in 86% yield by heating to reflux in toluene. To our delight, the protected substrate **28** indeed underwent Lacey–Dieckmann cyclization upon treatment with *t*BuOK (room temperature, 0.5 h) to give the corresponding tetramic acid **29** in 87% yield. Notably, the C14 and C15 proton signals of **29** appeared at higher field (C14,  $\delta$ =0.49; C15,  $\delta$ =0.80); this is ascribable to the anisotropic effect of the benzene ring within a rigid cyclophane skeleton.

The final step was the particularly problematic removal of the amide protecting group. After a number of unsuccessful attempts using other protecting groups, [24] the para-azidobenzyl protecting group,<sup>[22]</sup> employed in 29, was found to be the only group that could be successfully removed in the final step of the synthesis. Reduction of the azide moiety in 29 gave the corresponding amine, which, upon exposure to DDQ in the presence of water, afforded macrocidin A (1) as a pale beige solid that exhibited physical properties consistent with the reported data (1H and 13C NMR, IR, mass spectra).[1] Although the melting point was not reported in the original paper, [1] reprecipitation (dichloromethane/methanol) gave 1 as a fine, pale beige powder with a melting point of 205-207 °C. The sign and magnitude of the optical rotation concurred well with the reported values of natural macrocidin A:  $[\alpha]_D^{27} = +42$  (c = 0.18, methanol), lit.  $[\alpha]_D^{25} = +45$  (c = 0.35, methanol), thus establishing the absolute configuration of the natural product as shown (Scheme 8).

**Scheme 8.** Total synthesis of macrocidin A. a)  $H_2$ , 10% Pd/C, MeOH, THF, RT, 2 h. b) DDQ,  $H_2O$ , THF, room temperature, 0.5 h (78%, 2 steps). DDQ = 2,3-dichloro-5,6-dicyano-1,4-benzoquinone.

In summary, we have achieved the first total synthesis of macrocidin A. and have established the absolute configuration. Further work is ongoing to synthesize macrocidin B and other analogues of biological relevance.

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